

Total Synthesis of Phomactin A[†]

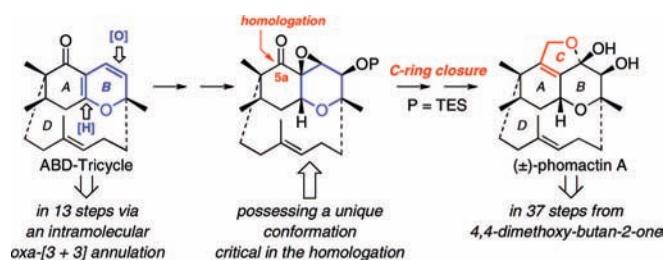
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ABSTRACT



A total synthesis of (±)-phomactin A is described to highlight the final completion of a complex natural product target that had commenced with an intramolecular oxa-[3 + 3] annulation strategy in the construction of the ABD-tricycle. These efforts reveal structural intricacies of this ABD-tricycle with an illustrative example being the conformational analysis that was ultimately critical for the C5a-homologation.

In the past decade, phomactin A,^{1–4} a structurally unique natural product isolated from the culture filtrate of a parasitic fungus *Phoma* sp. (SANK 11486) found on the shell of *Chinoecetes opilio*, has captured an impressive array of synthetic efforts.^{5,6} Although possessing only modest inhibition against platelet-activating factor⁷ (PAF)-induced platelet aggregation (IC₅₀ = 10 μM), phomactin A embodies a new class of PAF antagonists. The most active member is D and was synthesized by Yamada.⁸ Phomactin A is the only known tetracycle (discounting epoxides) in the phomactin family with all other members lacking either the B-ring or C-ring. Thus, phomactin A represents structurally the most complex member. To date, two elegant total syntheses of

(±)- and (+)-phomactin A were completed by Pattenden⁹ and Halcomb,¹⁰ respectively. Recently, Wulff¹¹ reported the synthesis of phomactin B2.

[†] This paper is dedicated to Professor Bill Wulff on the very special occasion of his 60th birthday.

(1) Sugano, M.; Sato, A.; Iijima, Y.; Oshima, T.; Furuya, K.; Kuwano, H.; Hata, T.; Hanzawa, H. *J. Am. Chem. Soc.* **1991**, *113*, 5463.

(2) (a) Sugano, M.; Sato, A.; Saito, K.; Takaiishi, S.; Matsushita, Y.; Iijima, Y. *J. Med. Chem.* **1996**, *39*, 5281. (b) Sugano, M.; Sato, A.; Iijima, Y.; Furuya, K.; Hata, T.; Kuwano, H. *J. Antibiot.* **1995**, *48*, 1188. (c) Sugano, M.; Sato, A.; Iijima, Y.; Furuya, K.; Haruyama, H.; Yoda, K.; Hata, T. *J. Org. Chem.* **1994**, *59*, 564.

(3) Koyama, K.; Ishino, K.; Takatori, K.; Takashi Sugita, T.; Kinoshita, K.; Takahashi, K. *Tetrahedron Lett.* **2004**, *45*, 6947.

(4) Also see: Zhu, X.; Lambertino, A. T.; Houghton, T. J.; McGilvra, J. D.; Xu, C.; Rawal, V. H.; Leff, A. R. *Life Sci.* **2003**, *73*, 3005.

(5) For reviews on synthetic efforts toward phomactins, see: (a) Cole, K. P.; Hsung, R. P. *ChemTracts* **2003**, *16*, 811. (b) Goldring, W. P. D.; Pattenden, G. *Acc. Chem. Res.* **2006**, *39*, 354.

(6) For synthetic approaches, see: (a) Foote, K. M.; Hayes, C. J.; Pattenden, G. *Tetrahedron Lett.* **1996**, *37*, 275. (b) Chen, D.; Wang, J.; Totah, N. I. *J. Org. Chem.* **1999**, *64*, 1776. (c) Seth, P. P.; Totah, N. I. *J. Org. Chem.* **1999**, *64*, 8750. (d) Seth, P. P.; Totah, N. I. *Org. Lett.* **2000**, *2*, 2507. (e) Kallan, N. C.; Halcomb, R. L. *Org. Lett.* **2000**, *2*, 2687. (f) Chemler, S. R.; Danishefsky, S. J. *Org. Lett.* **2000**, *2*, 2695. (g) Seth, P. P.; Chen, D.; Wang, J.; Gao, X.; Totah, N. I. *Org. Lett.* **2000**, *2*, 10185. (h) Foote, K.; John, M.; Pattenden, G. *Synlett* **2001**, 365. (i) Mi, B.; Maleczka, R. *Org. Lett.* **2001**, *3*, 1491. (j) Chemler, S. R.; Iserloh, U.; Danishefsky, S. J. *Org. Lett.* **2001**, *3*, 2949. (k) Houghton, T.; Choi, S.; Rawal, V. H. *Org. Lett.* **2001**, *3*, 3615. (l) Mohr, P. J.; Halcomb, R. L. *Org. Lett.* **2002**, *4*, 2413. (m) Balnaves, A. S.; McGowan, G.; Shapland, P. D. P.; Thomas, E. J. *Tetrahedron Lett.* **2003**, *44*, 2713. (n) Cheing, J. W. C.; Goldring, W. P. D.; Pattenden, G. *Chem. Commun.* **2003**, 2788. (o) Ryu, K.; Cho, Y.-S.; Jung, S.-I.; Cho, C.-G. *Org. Lett.* **2006**, *8*, 3343. (p) Huang, J.; Wang, H.; Wu, C.; Wulff, W. D. *Org. Lett.* **2007**, *9*, 2799. (q) Seth, P. P.; Chen, D.; Wang, J.; Gao, X.; Totah, N. I. *Tetrahedron Lett.* **2007**, *48*, 4605. (r) Peng, W.; Lee, C.-S. *Synlett* **2008**, 142. (s) DiBlasi, C. M.; Hamblett, C. L.; Leighton, J. L. *Abstracts of Papers*, 224th National Meeting of the American Chemical Society, Boston, MA, Fall, 2002; American Chemical Society: Washington, DC, 2002; ORGN-364.

(7) Platelet-activating factor is a phospholipid mediator released in the body by several cell types that play a role in causing inflammatory diseases. See: (a) Koltai, M.; Braquet, P. G. *Clin. Rev. Allergy* **1994**, *12*, 361. (b) Xhu, X.; Muñoz, N. M.; Kim, K. P.; Sano, H.; Cho, W.; Leff, A. R. *J. Immunol.* **1999**, *163*, 3423.

(8) For a total synthesis of (+)-phomactin D, see: Miyaoka, H.; Saka, Y.; Miura, S.; Yamada, Y. *Tetrahedron Lett.* **1996**, *37*, 7107.

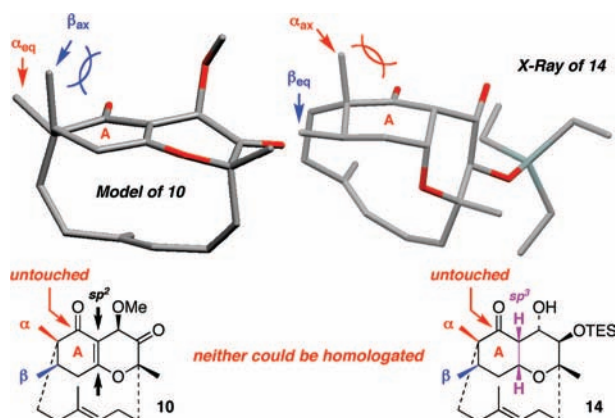
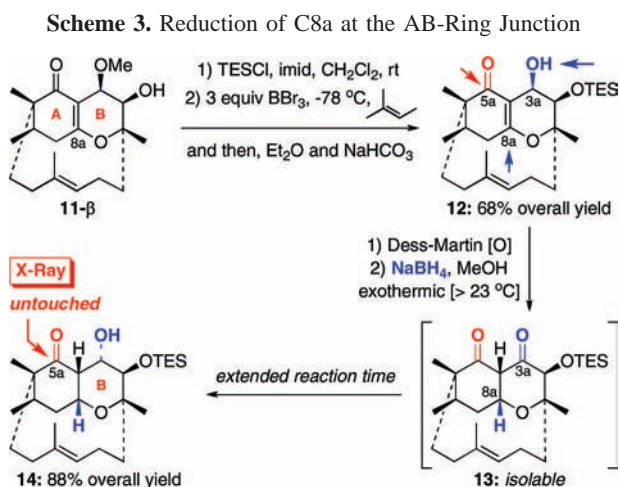


Figure 1. Conformational analysis for **10** and **14**.

the facial differentiation of the reduction. With a more bulky hydride, the C3a-OMe group was able to better prevent the hydride approaching from the top face.

Reduction at C8a was relatively less eventful. As shown in Scheme 3, capping of C3-OH in **11- β** with TESCl



followed by demethylation with BBr_3 led to vinylogous ester **12**. However, no condition that we screened (i.e., L-Selectride, Cu/LAH , or Na/IPA) was capable of reducing the vinylogous ester motif in a 1,4-manner. Realizing that vinylogous ester **12** may not be sufficiently electron deficient, we oxidized C3a-OH using Dess–Martin periodinane reagent, and an ensuing reduction effectively gave diketone **13**, which is isolable, but with extended reduction time at temperatures slightly greater than rt, hydroxy ketone **14** was obtained in 96% overall yield with completely selective reduction at C3a.

With hydroxy ketone **14**, we completed our efforts in transforming B-ring into its proper oxidation states, and a single-crystal X-ray structure of **14** (Figure 1, right side) further affirms our success. However, we were concerned

about the reactivity of the C5a carbonyl group because the challenge of C5a-homologation lies ahead. It became obvious that no reduction of the C5a carbonyl group in **10** had occurred when using L-Selectride, and nor did NaBH_4 touch the C5a carbonyl group in **13**. While one could concede that reduction in **10** involved a vinylogous ester, lack of reduction in **13** was quite disconcerting.

Upon examination of the minimized Spartan model of **10** and X-ray structure of **14**, we found some unique conformational elements. In **10**, the α -Me group in the A-ring (red) is pseudoequatorial with the β -Me group (blue) being pseudoaxial, thereby blocking any incoming nucleophiles toward the C5a carbonyl group. On the other hand, hydroxy ketone **14** assumes a very different conformation with its AB-ring junction being both sp^3 -hybridized instead of sp^2 as in **10**. In this case, the β -Me group (blue) is now pseudoequatorial with the α -Me group (red) turning to occupy the pseudoaxial position, thereby hindering the attack of the C5a carbonyl group.

In contrast, the AB-ring junction of endoperoxide **8** consists of sp^3 -hybridized C8a and sp^2 -hybridized C8b (Figure 2). This set of hybridizations leads to yet another

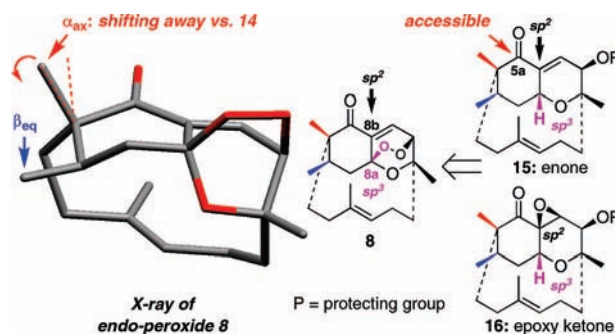


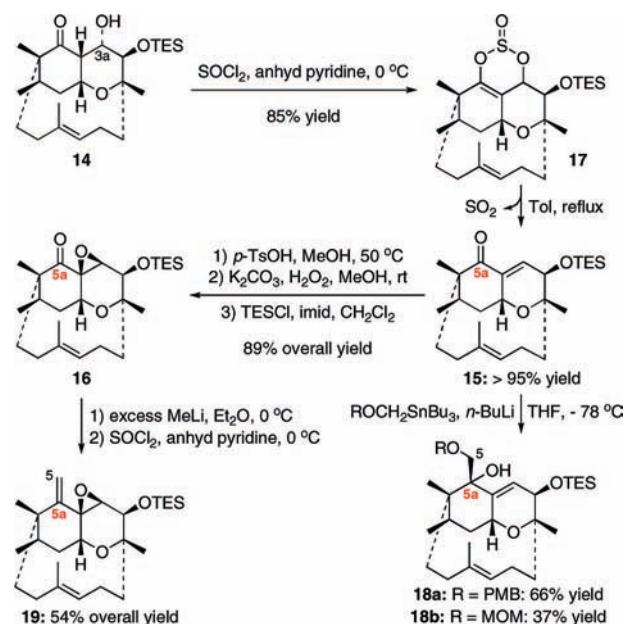
Figure 2. Rationale for choosing **15** and **16**.

conformation in which the β -Me group (blue) remains pseudoequatorial as in **14**, but the α -Me group (red) shifts away versus its respective position in **14**. We hoped that this minor shift would provide just enough opening to allow C5a to be accessible for homologation.

To test this hypothesis, we elected to construct enone **15** and epoxy ketone **16** for which both ring-junctions contain one sp^3 - and one sp^2 -hybridized carbon.²² As shown in Scheme 4, after failing to eliminate the C3a-OH group via dehydrative protocols, we isolated sulfite **17** during an attempt to chlorinate at C3a. A retro-Diels–Alder process in refluxing toluene would extrude SO_2 and afford the desired enone **15**. To our relief, we could add various one-carbon nucleophiles such as ROCH_2Li ($\text{R} = \text{PBMB}$ or MOM) to afford ene–diols **18a/b**, thereby succeeding what had appeared to be a daunting task in C5a-homologation. However, ene–diols **18a/b** were not useful for the total synthesis.

(22) Spartan models of enone **15** and epoxy ketone **16** reveal conformations similar to that of endo-peroxide **8**.

Scheme 4. C5a-Homologation of Enone **15** and Epoxy Ketone **16**

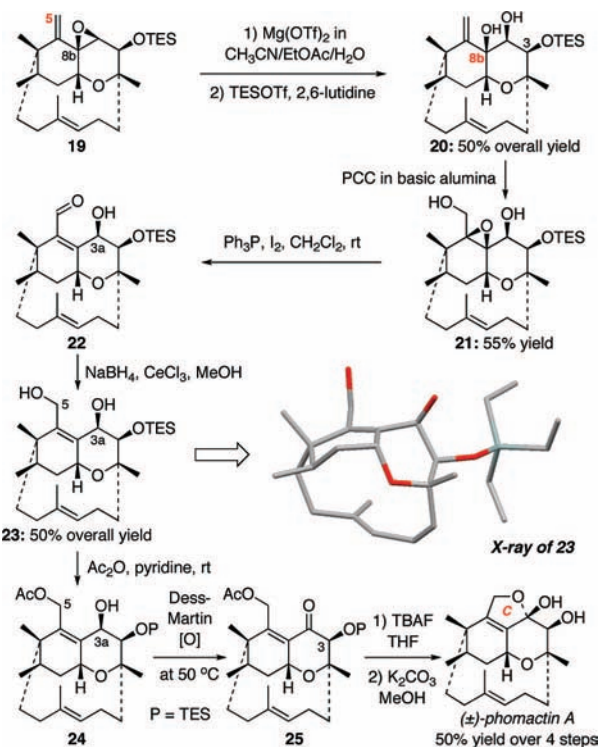


Consequently, we prepared epoxy ketone **16**, but in three steps, because epoxidation of enone **15** would not take place unless the TES group was removed. Homologation of **16** via addition of MeLi followed by elimination gave vinyl epoxide **19**.

Nucleophilic ring-opening of vinyl epoxide **19** at C5 via an S_N2' pathway would have been truly welcome at this stage (Scheme 5). Instead, an S_N1 -like process occurred with H_2O adding at C8b with retention of stereochemistry when using $Mg(OTf)_2$ in wet CH_3CN ,²³ leading to 1,2-diol **20** after resilylating C3-OH. Allylic alcohol transposition using Dauben's protocol²⁴ led to epoxy diol **21**.^{25,26} Subsequent treatment of **21** with Ph_3P-I_2 ²⁷ followed by Luche reduction of the enal intermediate **22** gave 1,4-diol **23**, which was confirmed through its X-ray structure.

After failing a number of approaches for constructing the C-ring using either enal **22** or 1,4-diol **23**, mainly due to

Scheme 5. Completing a Total Synthesis of (±)-Phomactin A



our inability to consistently oxidize the C3a-OH group, we managed to first acrylate C5-OH. Subsequently, we found that oxidation of the C3a-OH group employing Dess–Martin periodinane reagent at a warmer temperature gave enone **25**. An ensuing deprotection sequence allowed for the formation of the lactol C-ring and the final completion of our total synthesis of (±)-phomactin A in 24 steps from ABD-tricycle **2**.

We have described a total synthesis of (±)-phomactin A that highlights the final completion of a complex natural product target that had commenced with an intramolecular *oxa*-[3 + 3] annulation strategy in the construction of an ABD-tricycle. Our efforts reveal structural intricacies of this ABD-tricycle with an illustrative example being the conformational analysis that was ultimately critical for the C5a-homologation.

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Supporting Information Available: Experimental procedures as well as NMR spectra, characterizations, and X-ray structural files. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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(23) For related protocols, see: (a) Boyer, F.-D.; Hanna, I. *J. Org. Chem.* **2005**, *70*, 1077. (b) Iranpoor, N.; Shekariz, M.; Shiriny, F. *Synth. Commun.* **1998**, *28*, 347.

(24) Dauben, W. G.; Michno, D. M. *J. Org. Chem.* **1977**, *42*, 682.

(25) For a recent example, see: (a) Chai, Y.; McIntosh, M. C. *Tetrahedron Lett.* **2004**, *45*, 3269. Also see: (b) Sundararaman, P.; Herz, W. *J. Org. Chem.* **1977**, *42*, 813. (c) Chu, A.; Mander, L. N. *Tetrahedron Lett.* **1988**, *29*, 2727.

(26) To circumvent this epoxide formation, we also attempted $MeReO_3$ that was successful in a related 1,3-transposition; see: (a) John, M.; Hutchison, J. M.; Lindsay, H. A.; Dormi, S. S.; Jones, G. D.; Vicic, D. A.; McIntosh, M. C. *Org. Lett.* **2006**, *8*, 3663. Also see: (b) Jacob, J.; Espenson, J. H.; Jensen, J. H.; Gordon, M. S. *Organometallics* **1998**, *17*, 1835. (c) Morrill, C.; Grubbs, R. H. *J. Am. Chem. Soc.* **2005**, *127*, 2842.

(27) Wydra, H.; Paryzek, Z. *Tetrahedron Lett.* **1984**, *25*, 2601.